



# 4-Aminoquinolines as a Novel Class of NR1/2B Subtype Selective NMDA Receptor Antagonists

Emmanuel Pinard, a,\* Alexander Alanine, Anne Bourson, Bernd Büttelmann, Marie-Paule Heitz, Vincent Mutel, Ramanjit Gill, Gerhard Trube and René Wyler

<sup>a</sup>Pharma Division, Discovery Chemistry, F. Hoffmann-La Roche Ltd., CH-4070 Basel, Switzerland <sup>b</sup>Pharma Division, Preclinical CNS Research, F. Hoffmann-La Roche Ltd., CH-4070 Basel, Switzerland

Received 20 February 2002; revised 17 May 2002; accepted 17 June 2002

**Abstract**—Screening of the Roche compound library led to the identification of 4-aminoquinoline **4** as structurally novel NR1/2B subtype selective NMDA receptor antagonist. The SAR which was developed in this series resulted in the discovery of highly potent and in vivo active blockers. © 2002 Elsevier Science Ltd. All rights reserved.

Overactivation of NMDA receptors and the resulting calcium overload of neurones is considered to be the main contributor to neuronal cell death following acute cerebral ischemia. NMDA receptor antagonists have, thus, been shown to be potent neuroprotective agents in animal models of focal cerebral ischemia. 1,2

Native NMDA receptors exist as heteromeric assemblies containing NR1 subunits together with one or more of the four NR2 subunits (NR2A-D).<sup>3,4</sup> During the last decade, a number of NR1/2B subtype selective blockers<sup>5</sup> have been described such as ifenprodil 1,<sup>6</sup> CP-101,606 2,<sup>7</sup> and Ro-25–6981 3.<sup>8</sup> These compounds showed neuroprotective effect in vivo without inducing the side effects associated with many non-selective NMDA receptor antagonists.<sup>9–12</sup> In addition, recent studies demonstrated that ifenprodil 1 produces a state-dependent block of NMDA receptors.<sup>13</sup> This makes NR1/2B subtype selective blockers potentially attractive drugs for the treatment of neurodegenerative diseases such as stroke,<sup>2</sup> brain trauma,<sup>2</sup> pain<sup>12,14</sup> and also Parkinson disease.<sup>15</sup>

Strikingly, most NR1/2B subtype selective NMDA receptor antagonists described so far are structurally related to ifenprodil 1. Key elements of the proposed pharmacophoric model in this series of compounds

include two aromatics, one central and usually basic nitrogen and a phenolic hydroxyl group or a bioisosteric phenol replacement.<sup>16</sup>

As part of our program, we set out to discover potent, in vivo active NR1/2B suptype selective blockers structurally unrelated to ifenprodil 1. Toward this end, screening of the Roche compound library using tritiated Ro-25–6981 as the radioligand in the binding assay was performed. This screening campaign led to the identification of the 4-aminoquinoline 4.

<sup>\*</sup>Corresponding author. Tel.: +41-61-688-4388; fax: +41-61-688-8714; e-mail: emmanuel.pinard@roche.com

Route 1: 
$$A9$$
 a Route 1:  $A3$ ,  $A8$ ,  $A9$ :  $R_1 = 2$ -yl-1,  $2$ ,  $3$ -4-tetrahydro-naphtalene Yield Route 2:  $A8$   $A8$   $A9$ :  $A$ 

Scheme 1. (a) cat. TsOH, toluene,  $110^{\circ}$ C, Dean–Stark then Ph<sub>2</sub>O,  $260^{\circ}$ C, 32%; (b) POCl<sub>3</sub>,  $80^{\circ}$ C, 64%; (c) R<sub>1</sub>X (X = Br or I), nBuLi, THF,  $-20^{\circ}$ C to rt then H<sub>2</sub>O, I<sub>2</sub>, NaOH, rt, 30–80%; (d) 1,2,3,4-tetrahydroisoquinoline, toluene,  $110^{\circ}$ C, 84%; (e) RNH<sub>2</sub>, 150– $200^{\circ}$ C, 60–100%; (f) PDC, DMF, rt, 6%.

Using 4 as a lead compound, potent and in vivo active NR1/2B suptype selective NMDA receptor antagonists were designed.

### Chemistry

The main synthetic scheme for the preparation of 4-aminoquinolines is straighforward using established synthetic methods and is outlined in Scheme 1.

The 4-amino groups of compounds 4, 6–27, 29–32 were introduced by heating the corresponding 4-chloroquinolines 33–47 with an excess of primary amines in the absence of any solvent. Ketone 28 was produced by oxidation of aminopropan-2-ol derivative 22 (see Table 3) in the presence of PDC. Depending on the nature of the substituents located at position 2 of the quinoline ring, the non commercially available 4-chloroquinolines 33–44 were prepared according to three different routes. Following route one, 4-chloroquinoline 43 was produced by chlorination of the corresponding quinolone **48**<sup>17</sup> which was obtained by thermal condensation of aniline and β-ketoester 49.18 Following route two, 4-chloroquinolines 33-42 were obtained by adding alkyl or aryl lithium derivatives to 4-chloroquinoline followed by aromatisation of the intermediates 1,2-dihydroquinolines with iodine and base. 19,20 Finally, according to route three, 4-chloroquinoline 44 was prepared in a regioselective manner by condensation of 2,4-dichloroquinoline with 1,2,3,4-tetrahydroisoquinoline.

#### Results and Discussion

Our first aim was to define the essential structural elements required for in vitro activity.

As shown in Table 1, starting from the screening hit 4, activity was abolished by removing the aminopropandiol moiety (5) and recovered after introduction of the amino group at position 4 of the quinoline ring (6). This result demonstrated that the nitrogen atom at position 4 is crucial for activity. As expected, introduction of the nitrogen atom has a profound effect on the basicity of the quinoline ring (see Table 1). It renders it sufficiently basic so that it is protonated at physiological pH (pK > 8.4). This observation strongly suggests that the protonated quinoline is most likely to be the bioactive form which interacts with the NMDA receptor.

In addition, it was found that the phenyl group at position 2 of the quinoline system was also essential since its deletion or its replacement by aliphatic groups resulted in virtually inactive compounds (7–9).

Having identified these two important structural elements, the next step was to explore chemical variations of the 2-aryl ring and of the 4-amino group.

As shown in Table 2, introduction of a substituent in ortho position of the 2-aryl ring resulted in the weakly active compound 10, suggesting that coplanarity of the aromatic ring with the quinoline nucleus is required.

**Table 1.** NMDA affinities of compounds 4–9: influence of  $pK_a$ 

Compd	$R_1$	$R_2$	K <sub>i</sub> (nM) <sup>a</sup> NMDA <sup>b</sup>	pK <sub>a</sub> <sup>c</sup>
4		у ОН ОН	260	8.4
5		Н	> 35,000	5.2
6		$NH_2$	413	8.5
7	Н	N OH OH	> 35,000	n.d.
8	CH <sub>3</sub>	У ОН ОН	>40,000	n.d.
9	(CH <sub>2</sub> ) <sub>3</sub> CH <sub>3</sub>	N OH OH	33,000	n.d.

<sup>&</sup>lt;sup>a</sup>Binding affinities are quoted as  $K_i$  values and are the geometric mean of at least two experiments. The variability is less than 20%.

Furthermore, introduction of small and lipophilic substituents in the *para* position (11–13) showed a beneficial effect. In particular, highest activity was obtained in the case of the presence of electron donating substituents. Introduction of larger (14) or more polar

substituents (15) at the para position led to a significant drop of affinity. This result strongly suggests that the aromatic moiety fits in a size limited hydrophobic pocket of the NMDA receptor. Compounds substituted at both the *meta* and *para* position by small and lipophilic substituents generally showed high activity as exemplified with compounds 16 and 17. Low nanomolar activity was also obtained for the annelated derivative Interestingly, the regioisomeric tetrahydronaphtalene derivative 19 was found to be fairly active (100 nM) demonstrating that the aromatic group does not need to be directly attached at position 2 of the quinoline ring. Gratifyingly, a 5-fold increase of activity was observed by attaching the quinoline ring by the means of a C-N bond instead of a C-C bond as illustrated by the tetrahydroisoquinoline derivative **20** (11 nM).

By keeping the phenyl group fixed at  $C_2$ , variation of the  $C_4$  position was explored on the quinoline ring system.

As exemplified in Table 3, starting from the lead compound 4, deletion of both OH groups resulted in nearly 20-fold decrease of activity (21) demonstrating that at least one OH group is mandatory for good binding affinity. The fact that activity was retained with the racemic 2-amino propanol derivative 22 and dramatically reduced with the aminopropanol derivative 23 demonstrated that the important OH group is the one located at the vicinal position to the nitrogen atom. Of the two possible enantiomers of 22, the *R*-configurated 2-aminopropanol derivative 24 displayed the best affinity for the NMDA receptor (75 nM). The same range of activity was also attained with the non-chiral ethanolamino derivative 26. As illustrated by compounds 27

Table 2. NMDA affinities of compounds 10-20; variation of R<sub>1</sub>

Compd	$R_1$	K <sub>i</sub> (nM) <sup>a</sup> NMDA <sup>b</sup>	Compd	$R_1$	K <sub>i</sub> (nM) <sup>a</sup> NMDA <sup>b</sup>
10		3700	16	CI	17
11		23	17		26
12		19	18		22
13	a	100	19		100
14	Col	500	20		11
15	ОН	> 4000			

<sup>&</sup>lt;sup>a</sup>See Table 1.

<sup>&</sup>lt;sup>b</sup>Displacement of [<sup>3</sup>H]-25–6981.<sup>21</sup>

 $<sup>{}^{</sup>c}pK_{a}$  values were determined using a potentiometric method.<sup>22</sup>

<sup>&</sup>lt;sup>b</sup>See Table 1.

Table 3. NMDA affinities of compounds 21–28; variation of R<sub>2</sub>

Comp	$R_2$	K <sub>i</sub> (nM) <sup>a</sup> NMDA <sup>b</sup>	Compd	$R_2$	K <sub>i</sub> (nM) <sup>a</sup> NMDA <sup>b</sup>
21	× <sub>H</sub> ~	4900	25	HOH	450
22	N OH	260	26	, он Н	110
23	, <sup>Н</sup> ОН	1900	27	, H	7500
24	N OH	75	28	K N N	7900

<sup>&</sup>lt;sup>a</sup>See Table 1.

Table 4. NMDA affinities and in vivo activities of compounds 29–32

Compd	R <sub>1</sub>	$R_2$	K <sub>i</sub> (nM) <sup>a</sup> NMDA <sup>b</sup>	ED <sub>50</sub> (mg/kg) <sup>c</sup> Sound induced seizures
29		, Å OH	3.5	<12
30	CI	A OH	10	12
31		N OH	10	17
32		, NOH	15	25

<sup>&</sup>lt;sup>a</sup>See Table 1.

and 28, the potency was severely reduced after methylation or oxidation of the vicinal OH group, suggesting that this group acts as an hydrogen bond donor.

The next step in our optimization phase was to combine the best substituents which were previously identified at position 2 and 4. As exemplified in Table 4, the effect of substituents was found to be additive and compounds with much improved in vitro activity were identified. In particular, exceptionally high potency (3.5 nM) was achieved with the tetrahydroisoquinoline derivative 29 containing the ethanolamino group at the C<sub>4</sub> position.

In vivo activity was measured in mice after ip administration<sup>23</sup> using the standard sound-induced seizures assay.<sup>24</sup> As depicted in Table 4, the 4-aminoquinoline

derivatives exhibited good in vivo activity indicating that they penetrate well into the brain.

#### Conclusion

Starting from 4-aminoquinoline 4 as a screening hit, a SAR was developed which resulted in the discovery of highly in vitro and in vivo potent NR1/2B subtype selective NMDA antagonists structurally unrelated to ifenprodil. Within this novel series, the pharmacophore elements identified were: (1) a protonated quinoline ring, (2) a lipophilic aromatic system attached at the C<sub>2</sub> position and (3) a hydrogen bond donating group linked vicinally to a C<sub>4</sub>-nitrogen on the quinoline framework. Since the nature and the spatial arrangement

<sup>&</sup>lt;sup>b</sup>See Table 1.

<sup>&</sup>lt;sup>b</sup>See Table 1.

<sup>&</sup>lt;sup>c</sup>Compounds were administered ip 30 min before testing in DBA/2 mice.

of these pharmacophore elements differ notably from those identified in the ifenprodil series, it is suggested that the 4-aminoquinolines reported here and the ifenprodil like compounds might not fully share the same binding site at the NMDA receptor.

## Acknowledgements

The skillful technical assistance of S. Burner, T. Muser, and J. Padilla is gratefully acknowledged. The authors wish to thank W. Arnold, W. Meister for spectroscopic characterization and Bjorn Wagner for  $pK_a$  determinations.

#### References and Notes

- 1. Kemp, J. A.; Kew, J. N. C.; Gill, R. In *Handbook of Experimental Pharmacology*; Jonas, P., Monyer, H., Eds.; Springer: Berlin, Heidelberg, 1999, Vol. 141, p 495.
- 2. Gill, R.; Kemp, J. A.; Richards, J. G.; Kew, J. N. C. Curr. Opin. Cardiovasc., Pulm. Renal Invest. Drugs 1999, 1, 576.
- 3. Monyer, H.; Sprengel, R.; Schoepfer, R.; Herb, A.; Higuchi, M.; Lomeli, H.; Burnashev, N.; Sakmann, B.; Seeburg, P. H. *Science* **1992**, *256*, 1217.
- 4. Moriyoshi, K. M.; Masu, M.; Ishii, T.; Shigemoto, R.; Mizuno, N.; Nakanishi, S. *Nature (London)* **1991**, *354*, 31.
- 5. Chenard, B. L.; Menniti, F. S. Curr. Pharm. Des. 1999, 5, 381.
- 6. Williams, K. Mol. Pharmacol. 1993, 44, 851.
- 7. Chenard, B. L.; Bordner, J.; Butler, T. W.; Chambers, L. K.; Collins, M. A.; De Costa, D. L.; Ducat, M. F.; Dumont, M. L.; Fox, C. B.; Mena, E. E.; Meniti, F. S.; Nielsen, J.; Pagnozzi, M. J.; Richter, K. E. G.; Ronau, R. T.; Shalaby, I. A.; Stemple, J. Z.; White, W. F. *J. Med. Chem.* 1995, 38, 3138.
- 8. Fischer, G.; Mutel, V.; Trube, G.; Malherbe, P.; Kew, J. N. C.; Mohacsi, E.; Heitz, M. P.; Kemp, J. A. *J. Pharmacol. Exp. Ther.* **1997**, *283*, 1285.

- 9. Shalaby, I. A.; Chenard, B. L.; Prochniak, M. A.; Butler, T. W. J. Pharmacol. Exp. Ther. 1992, 260, 925.
- 10. Menniti, F.; Chenard, B.; Collins, M.; Ducat, M.; Shalaby, I.; White, F. Eur. J. Pharmacol. 1997, 331, 117.
- 11. Fischer, G.; Bourson, A.; Kemp, J. A.; Lorez, H. P. Neuroscience Annual Meeting, Washington, DC, Nov 16–21, 1996.
- 12. Boyce, S.; Wyatt, A.; Webb, J. K.; O'Donnell, R.; Mason, G.; Rigby, M.; Sirinathsinghji, D.; Hill, R. G.; Rupniak, N. M. J. *Neuropharm.* **1999**, *38*, 611.
- 13. Kew, J. N. C.; Trube, G.; Kemp, J. A. J. Phys. 1996, 497, 761
- 14. Taniguchi, K.; Shinjo, K.; Mizutani, M.; Shimada, K.; Ishikawa, O.; Menniti, F. S.; Nagahisa, A. *Br. J. Pharmacol.* **1997**, *122*, 809.
- 15. Wright, J. L.; Gregory, T. F.; Boxer, P. A.; Meltzer, L. T.; Serpa, K. A.; Wise, L. D. *Bioorg. Med. Chem. Lett.* **1999**, *9*, 2815.
- 16. Tamiz, A. P.; Whittemore, E. R.; Zhou, Z.-L.; Huang, J.-C.; Drewe, J. A.; Chen, J.-C.; Cai, S.-X.; Weber, E.; Woodward, R. M.; Keana, J. F. W. *J. Med. Chem.* **1998**, *41*, 3499.
- 17. Alanine, A.; Burner, S.; Buettelmann, B.; Heitz Neidhart, M.-P.; Jaeschke, G.; Pinard, E.; Wyler, R. Eur. Pat. Appl. EP 1088818, 2001. *Chem. Abstr.* **2001**, *134*, 280718.
- 18. Hauser, C. R.; Reynolds, G. A. J. Am. Chem. Soc. 1948, 70, 2402.
- 19. Gilman, H.; Benkeser, R. A. J. Am. Chem. Soc. 1947, 69, 123
- 20. Field, G. F.; Zally, W. J. US Patent 4,560,692, 1985. *Chem. Abstr.* **1985**, *105*, 97339.
- 21. Mutel, V.; Buchy, D.; Klingelschmidt, A.; Messer, J.; Bleuel, Z.; Kemp, J. A. *J. Neurochem.* **1998**, *70*, 2147.
- 22. Avdeef, A. Applications and Theory Guide to pH-Metric  $pK_a$  and logP Measurement; Sirius Analytical Instruments: Forest Row, UK, 1993.
- 23. Activity after oral administration was not determined.
- 24. Bourson, A.; Kapps, V.; Zwingelstein, C.; Rudler, A.; Boess, F. G.; Sleight, A. J. *Naunyn-Schmiedeberg's Arch. Pharmacol.* **1997**, *356*, 820.